INTENDED USE

The ZENIT RA CARDIOLIPIN IgG test is a chemiluminescent immunoassay (CLIA) for use on the dedicated ZENIT RA Analyzer for quantitative determination of the specific β₂-glycoprotein I dependent anti-cardiolipin IgG antibodies in samples of human serum or plasma (EDTA, heparin).

This assay method is employed as a supplementary diagnostic technique in evaluation of antiphospholipid syndrome (APS).

CAUTION: Medical decisions cannot be based solely on the results of this test but must take into account all available clinical and laboratory data.

CLINICAL SIGNIFICANCE

The presence of antiphospholipid (aPL) antibodies in patients with venous and/or arterial thrombosis or in patients with pregnancy-related complications is the essential laboratory marker (together with LAC [lupus anticoagulant] testing) for diagnosis of antiphospholipid antibody syndrome (or antiphospholipid syndrome) (APS or APLS).¹

In accordance with the Sapporo criteria, updated in 2006,¹ APS can be definitively diagnosed in the presence of at least one clinical criterion and one laboratory criterion.

The laboratory criteria are persistent positivity (12 weeks) with an average/high titer for anti-cardiolipin (aCL) and/or anti-β₂-glycoprotein I (a-β₂GPI) antibodies and/or “lupus anticoagulant” (LAC) antibodies. The aCL and a-β₂GPI antibody isotype may be G or M and the antibodies may be present in titers in excess of 40 U/ml.

Antiphospholipid antibodies were first noted in 1941 in samples of patients with serologic diagnoses of syphilis.² It was shown that the serum of these patients inter-reacted with the cardiolipin phospholipid contained in the beef heart extract used in the VDRL (Venereal Disease Research Laboratory) test, which is considered specific for diagnosis of syphilis.

The specificity of the VDRL assay was challenged by the numerous false positive results in samples from patients with systemic autoimmune diseases in the absence of venereal diseases. In 1983, Harris et al.,³ applying a highly-sensitive method for detection of anticardiolipin antibodies, found high concentrations of aCL in 61% of patients with SLE (systemic lupus erythematosus), thus demonstrating a significant correlation between antibody levels and venous and arterial thrombosis, “lupus anticoagulant,” and thrombocytopenia.
In 1990, two independent groups of researchers demonstrated that the presence of \( \beta_2 \)-glycoprotein I is indispensable for detecting anticardiolipin antibodies.

\( \beta_2 \)-glycoprotein I has a molecular weight of 50 kDa ca., a plasma concentration of 0.15-0.30 mg/ml ca., and a biological function that is still not fully understood (although it would seem that it can modulate lipoprotein metabolism, interfere with some coagulation reactions, and act as a platelet anti-aggregant\(^6\)). Recent crystallographic studies have defined the three-dimensional structure of the protein and its 5-domain organization,\(^10\)\(^-\)\(^11\) providing useful information about the way this molecule works.

In detail, the fifth domain shows numerous lysine residues, which are responsible for the electrostatic interaction of \( \beta_2 \)-glycoprotein I with the anionic phospholipids of the cell membranes.\(^12\) The same mechanism is responsible for in vitro bonding between \( \beta_2 \)-glycoprotein and cardiolipin adsorbed to a solid phase. It has been amply demonstrated that the anticardiolipin antibodies of patients affected with antiphospholipid antibody syndrome recognize a modified portion of the \( \beta_2 \)-glycoprotein I molecule; these autoantibodies cannot recognize cardiolipin or native \( \beta_2 \)-glycoprotein not bound to solid phases or to other structures.\(^4\),\(^5\),\(^13\)\(^-\)\(^15\)

Knowledge acquired to date permits us to define the anticardiolipin antibodies as antibodies that can bind to neoepitopes generated by binding of \( \beta_2 \)-glycoprotein and cardiolipin adsorbed to a solid phase.

It was later shown that the anticardiolipin antibodies in patients with autoimmune diseases can recognize the \( \beta_2 \)-glycoprotein I directly adsorbed on UV-treated or irradiated polystyrene microtiter plates. In this case as well, recognition of the molecule by the autoantibodies is determined by the structural modifications caused when the protein binds to the solid phase.

**PRINCIPLE OF THE METHOD**

The ZENIT RA CARDIOLIPIN IgG kit for quantitative determination of the specific \( \beta_2 \)-glycoprotein I dependent anti-cardiolipin IgG antibodies employs an indirect, two-step immunoassay method based on the principle of chemiluminescence.

Cardiolipin/\( \beta_2 \)-glycoprotein I complex is used to coat magnetic particles (solid phase) and a human anti-IgG antibody is labeled with an acridine ester derivative (conjugate).

During the first incubation, the specific antibodies present in the sample, in the calibrators, or in the controls bind with the solid phase.

During the second incubation, the conjugate reacts with the anti-cardiolipin IgG antibodies captured on the solid phase.

After each incubation, the material that has not bonded with the solid phase is removed by suction and repeated washing.

The quantity of marked conjugate bonded to the solid phase is evaluated by chemiluminescent reaction and measurement of the light signal. The generated signal, measured in RLU (Relative Light Units), is indicative of the concentration of the specific antibodies present in the sample, in the calibrators, and in the controls.
AUTOMATION

The ZENIT RA Analyzer automatically performs all the operations called for by the assay protocol: addition of the samples, calibrators, controls, magnetic particles, conjugate, and chemiluminescence activator solutions to the reaction vessel; magnetic separation and washing of the particles; measurement of the emitted light.

The system calculates the assay results for the samples and the controls using the stored calibration curves and prints a report containing all the assay- and patient-related information.

MATERIALS AND REAGENTS

Materials and Reagents Provided

<table>
<thead>
<tr>
<th>REAG</th>
<th>1</th>
<th>MP</th>
<th>2.5 ml</th>
</tr>
</thead>
</table>

Magnetic particles coated with cardiolipin/β2-glycoprotein I complex in phosphate buffer containing stabilizing proteins, detergent, and Pro-Clin 300 and sodium azide (< 0.1%) as preservatives.

<table>
<thead>
<tr>
<th>REAG</th>
<th>2</th>
<th>CONJ</th>
<th>25 ml</th>
</tr>
</thead>
</table>

Mouse monoclonal anti-human IgG antibody labeled with an acridine ester derivative (conjugate), in phosphate buffer containing stabilizer proteins and sodium azide (< 0.1%) as preservative.

<table>
<thead>
<tr>
<th>REAG</th>
<th>3</th>
<th>DIL</th>
<th>25 ml</th>
</tr>
</thead>
</table>

Sample Dilution Solution: phosphate buffer containing bovine serum albumin, detergent, a blue dye, and Pro-Clin 300 and Gentamicin SO4 as preservatives.

<table>
<thead>
<tr>
<th>REAG</th>
<th>4</th>
<th>CAL A</th>
<th>1.6 ml</th>
</tr>
</thead>
</table>

Human serum with a low concentration of anti-cardiolipin IgG antibodies in phosphate buffer containing bovine serum albumin, detergent, inert blue dye, and Pro-Clin 300 and Gentamicin SO4 as preservatives.

<table>
<thead>
<tr>
<th>REAG</th>
<th>5</th>
<th>CAL B</th>
<th>1.6 ml</th>
</tr>
</thead>
</table>

Human serum with a high concentration of anti-cardiolipin IgG antibodies in phosphate buffer containing bovine serum albumin, detergent, inert blue dye, and Pro-Clin 300 and Gentamicin SO4 as preservatives.

All the reagents are ready to use.
Reagents 1, 2, and 3 are assembled into a single reagents cartridge unit.
The concentrations of the calibrators are expressed in GPL U/ml (U = units) and calibrated against the “Harris” reference sera. The concentration values are specific by product lot and are recorded on the DATA DISK included in the kit.

A mini-DVD containing information about all the ZENIT RA products (Reagents, Calibrators, Control Sera), updated to the latest production lot and excluding products expired at the date of writing of each new DATA DISK.

The only DATA DISK needed to ensure that the information needed for correct system operation is always updated is that with the highest lot number.

**Materials and Reagents Required but not Provided in the Kit**

- ZENIT RA Analyzer
  Code No. 41400
- ZENIT RA Cuvette Cube *
  Code No. 41402
  Box of 960 cuvettes.
- ZENIT RA System Liquid *
  Code No. 41409
  1 – 0.5-liter bottle of 10x solution.
- ZENIT RA Wash Solution *
  Code No. 41407
  1 – 0.5-liter bottle of 20x solution.
- ZENIT RA Trigger Set *
  Code No. 41403
  1 – 250-ml vial of Trigger A (pre-activation solution)
  1 – 250-ml vial of Trigger B (activation solution)
- ZENIT RA D-SORB Solution
  Code No. 41436
  Box containing 2 – 1-liter bottles of ready-to-use solution.
- ZENIT RA Cartridge Checking System *
  Code No. 41401
- ZENIT RA Top Cap Set
  Code No. 41566
  300 top caps for capping the calibrator containers after first use.

(*)The ZENIT RA Analyzer and the accessories tagged with an asterisk are manufactured by Immunodiagnostic Systems S.A., Rue E. Solvay, 101, B-4000 Liège, Belgium, and distributed by A. Menarini Diagnostics Srl.
Other Recommended Reagents

ZENIT RA APS IgG CONTROL SET

Code No. 41450

3 – 1.5 ml vials of human serum negative for anti-cardiolipin antibodies and 3 – 1.5 ml vials of human serum positive for anti-cardiolipin antibodies.

WARNINGS AND PRECAUTIONS

The reagents provided in the ZENIT RA CARDIOLIPIN IgG kit are intended for in vitro diagnostic use only and not for in vivo use in humans or animals.

This product must be used by professional users only and in strict accordance with the instructions set out in this document.

Menarini may not be held responsible for any loss or damage in any way resulting from or related to use of the product in manners not compliant with the instructions provided.

Safety Precautions

This product contains material of animal origin and therefore must be handled as though it contained infectious agents.

This product contains components of human origin. All the serum or plasma units used in the manufacture of the reagents in this kit have been tested by FDA-approved methods and have been found to be non-reactive for HBsAg and anti-HCV, anti-HIV1, and anti-HIV2 antibodies.

Nevertheless, since no analysis method can offer complete assurance of the total absence of pathogenic agents, all material of human origin should be considered potentially infected/infectious and be handled as such.

If the packaging is damaged in such a way as to cause leakage of the reagents, decontaminate the affected area with a dilute sodium hypochlorite (bleach) solution while wearing appropriate personal protective equipment (lab coat, gloves, goggles).

Dispose of used cleaning materials and the packaging materials affected by the leakage in accordance with national-level regulations for disposal of potentially infected/infectious waste.

Some reagents contain sodium azide as preservative. Since sodium azide may react with lead, copper, or brass in plumbing to form explosive azide compounds, do not flush reagents or waste to sewer. Dispose of such waste in accordance with national-level regulations for disposal of potentially hazardous substances.

Operating Precautions

In order to obtain reliable results, follow these instructions for use and the instructions provided in the analyzer operator’s manual carefully.

The reagents supplied in the kit are intended for use only with the ZENIT RA Analyzer system.

The reagents cartridge components cannot be removed from the cartridge and reassembled.
Do not use the kit after the expiry date.

REAGENT PREPARATION

The reagents supplied in the kit are ready to use.

REAGENT STORAGE AND STABILITY

Store the reagents supplied in the kit in an upright position, at 2-8 °C, in a dark place.
In these conditions, the reagents cartridges and the unopened calibrators reagents are stable until the expiry date.
After opening, the reagents cartridges may be used for 60 days if stored refrigerated at 2-8 °C or onboard the machine.
After opening, the calibrators may be used for 60 days if stored refrigerated at 2-8 °C or if the on-board use time does not exceed 6 hours per session.
Do not freeze the reagents and/or the calibrators.

SAMPLE PREPARATION AND STORAGE

The assay must be performed on samples of human serum or plasma (EDTA - heparin).
Use of lipemic, hemolyzed, or turbid samples is not recommended.
If the assay is performed more than 8 hours after the blood sample is drawn, after separating the serum from the coagulate or the plasma from the red blood cells transfer the surmatant from the gel separation tubes to secondary tubes.
Prior to analysis, the samples may be stored refrigerated at 2-8 °C for a maximum of 7 days.
If the samples must be stored for more than 7 days before testing, store frozen at < -20 °C.
Avoid repeated freezing and thawing.

ASSAY PROCEDURE

In order to obtain reliable analysis results, follow the instructions provided in the analyzer operator's manual carefully.

*Loading the Reagents*

All the reagents supplied in the kit are ready to use.
Before installing the reagents cartridge on the system, agitate the magnetic particles container by rotating horizontally, in order to ensure correct particle suspension. Do not allow the suspension to foam during agitation.

Position the reagents cartridge in the reagents area of the analyzer, using the guide for that purpose, and allow to agitate for at least 30 minutes prior to use.

The identification bar code is automatically read when the reagents cartridge is positioned on the analyzer. If the cartridge label is damaged or if for any other reason reading is not performed, the cartridge identification data may be entered manually.

The analyzer automatically keeps the magnetic particles suspension under agitation.

If the reagents cartridge is removed from the analyzer, store in an upright position, at 2-8 °C, in a dark place.

**Loading the Calibrators and Controls**

The ZENIT RA calibrators and controls are ready to use. Allow the calibrators and controls to stand at room temperature for 10 minutes before use. Agitate the contents gently, by hand or vortex; do not allow to foam. Do not upend the container and do not remove the seal cap with perforator (yellow cap for calibrators; green or blue caps for controls).

When using a calibrator or control for the first time, press the perforator cap down until it stops. This operation perforates the container seal membrane to permit accessing the liquid contents. If the perforator cap is used correctly, red strip at the top of the label will be covered (See Fig. 1 – Sealed Container and Perforated Container).

Previously-used calibrator and/or control containers will be capped with a top cap (white cap) and the red label strip will be covered.

Load only perforated containers from which the top cap (white cap) has been removed and on which the red strip is covered (Fig. 1 – Perforated Container) onto the analyzer.

Read the barcode and insert the calibrators or controls into the samples area of the analyzer. The barcode data may be entered manually if the label is damaged or if for any other reason reading is not performed.

The concentration values of the anti-cardiolipin IgG antibodies contained in the calibrators and the controls are stored on the DATA DISK and are automatically transferred to the analyzer. The data may be entered manually if for any reason data transfer is not successful.

At the end of each session, reseal the calibrator and control containers with the appropriate top caps (white caps) and transfer to storage at 2-8 °C until next use (See Fig. 1 – Capped Container).

The calibrators are sufficient for up to four uses.
Loading the Samples
Identify the samples via the barcode reader and insert them in the appropriate container on the analyzer. If a sample barcode is missing or illegible or for any other reason not read, the sample identification data may be entered manually.
Select the analysis parameters for each sample.

Calibration
The ZENIT RA Analyzer uses a calibration curve (master curve) that is generated by the manufacturer for each lot of reagents cartridges.
The master curve parameters, as well as the calibrator concentration values, are stored on the DATA DISK and transferred to the analyzer database.
Calibrators A and B are used for recalibrating the master curve for the instrument used and for the reagents installed onboard.
To recalibrate, analyze three replicates of the two calibrators (A and B) and one replicate of each control. The concentration values obtained with the controls permit validating the new calibration.
Once a master curve recalibration has been accepted and stored in memory, all the successive samples can be analyzed with no further calibration being required, exception made for the cases listed below:
- when a reagents cartridge with a new lot number is installed on the analyzer;
- when the control values do not fall within the acceptability interval;
- after analyzer maintenance;
- after expiry of the period of validity of the recalibrated master curve.
A master curve recalibrated for the ZENIT RA CARDIOLIPIN IgG kit has a period of validity of 15 days. Recalibration management is handled automatically by the analyzer.
**Assay**

Press the start button.

1. The system draws 100 µl of sample dilution solution, 20 µl of magnetic particles, 100 µl of sample dilution solution, and 6 µl of sample or control, in that order; for the calibrators, the positive serum is supplied prediluted with the sample dilution solution and the volume drawn is 106 µl. The solutions and the suspension are dispensed into the reaction cuvette.

2. The reaction cuvette is incubated on the rotor at 37°C for 10 minutes.

3. At the end of this incubation phase, the magnetic particles are separated and washed.

4. 200 µl of conjugate are dispensed into the cuvette.

5. The reaction cuvette is incubated on the rotor at 37°C for 10 minutes.

6. At the end of this last incubation phase, the magnetic particles are separated and washed and the cuvette is transferred to the reading chamber.

7. The quantity of conjugate bound to the solid phase, expressed in RLU (Relative Light Units), is directly proportional to the concentration of anti-cardiolipin IgG in the sample.

8. The results are interpolated on the calibration curve and expressed in concentrations.

If the concentration value of a sample exceeds the upper limit of the measurable interval, the sample may be diluted and retested. The new value thus obtained is multiplied by the appropriate dilution factor to obtain the final result.

**QUALITY CONTROL**

In order to ensure the validity of the assay, control sera at different concentrations (at least one negative serum and one positive serum) must be tested every day on which samples are assayed.

If individual laboratory practice so dictates, more frequent or more numerous controls may be performed for verification of assay results. Follow local quality control procedures.

If the ZENIT RA control sera are used, the expected mean values and the acceptability limits are those reported in the DATA DISK supplied with the controls.

Should different control sera be used, the expected values must be defined with the ZENIT RA reagents and analysis system before the products are used.

Should the values obtained with the controls not fall within the specified acceptability range, the relative assay results cannot be considered valid and it will be necessary to retest the respective samples.

In this case, recalibrate before repeating the assay/s in question.
CALCULATION AND INTERPRETATION OF RESULTS

Calculation of Results
The system automatically calculates the concentration of the anti-cardiolipin IgG antibodies in the tested sample. The values may be displayed on video or may be printed. The concentrations are expressed in GPL U/ml.
Calculation of the analyte concentration in a sample proceeds by reading of the result obtained for each sample on a calibration curve calculated in accordance with a 4-parameter logistic fitting model (4PL, weighted Y), which is corrected periodically on the basis of the calibrator assay results.

For detailed information on how the system calculates the results, refer to the system operator’s manual.

Interpretation of Results
The measurability range for the ZENIT RA CARDIOLIPIN IgG assay is: 0.0 – 640 GPL U/ml.
Values less than 0.0 GPL U/ml are extrapolated values and may be reported as “equal to 0.0 GPL U/ml.”
Values in excess of 640 GPL U/ml may be reported as “greater than 640 GPL U/ml” or the sample may be retested following appropriate dilution.
The results for a sample may be interpreted as set forth below:

<table>
<thead>
<tr>
<th>(GPL U/ml)</th>
<th>Interpretation</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt; 10</td>
<td>The sample should be considered Negative for the presence of anti-cardiolipin IgG.</td>
</tr>
<tr>
<td>10÷20</td>
<td>The sample should be considered inconclusive for the presence of anti-cardiolipin IgG.</td>
</tr>
<tr>
<td>&gt; 20</td>
<td>The sample should be considered Positive for the presence of anti-cardiolipin IgG.</td>
</tr>
</tbody>
</table>

The values reported above are suggested values only. Each laboratory will establish its own reference intervals.

LIMITS TO THE ASSAY METHOD

For diagnostic purposes, the results obtained with the ZENIT RA CARDIOLIPIN IgG kit and the ZENIT RA Analyzer system should always be used in conjunction with the other clinical and laboratory data available to the case physician.
Bacterial contamination of the samples and inactivation by heat may influence the results of the assay.
Heterophilic antibodies present in the human serum samples may react with immunoglobulin-based reagents, causing interference with in vitro immunoassays. Such samples may yield anomalous values when analyzed with the ZENIT RA CARDIOLIPIN IgG kit.
Samples from patients affected with chronic liver diseases, chronic infections, connective tissue disorders, and myeloma with hypergammaglobulinemia (IgG concentration in excess of 1800 mg/dl) may, in some cases, present positive values for anti-cardiolipin.

EXPECTED VALUES

Samples from 100 healthy subjects were analyzed to check for the presence of anti-cardiolipin IgG antibodies.
The results for all the samples analyzed were negative, with a mean value of 2.5 GPL U/ml and a standard deviation of 2.14 GPL U/ml.
The results thus obtained were used to calculate the “Limit of Blank” (LoB = the highest value that may be expected in a series of samples that do not contain the analyte). The Limit of Blank corresponding to the 95th percentile of the negative population was 6.2 GPL U/ml with reagents lot no. 2.

CLINICAL SENSITIVITY AND SPECIFICITY

A total of 244 samples were tested with the ZENIT RA CARDIOLIPIN IgG kit. Of these, 68 samples were from patients affected with antiphospholipid antibody syndrome (APS), 46 samples were from patients affected with systemic autoimmune rheumatic disorders (7 with connectivitis, 15 with systemic lupus erythematosus, 24 with rheumatoid arthritis), 30 samples were from patients affected with various infectious diseases (5 HIV, 7 HBV, 18 HCV), and 100 samples were from normal subjects.
In the presumably negative study population (46 samples from patients affected with systemic autoimmune rheumatic disorders, 30 samples from patients affected with various infectious diseases, and 100 samples from normal subjects), 5 samples tested positive, 6 tested inconclusive, and 165 tested negative.
- **Diagnostic Specificity:** 93.7 % (165/176)
In the presumably positive study population (68 samples from patients affected with antiphospholipid antibody syndrome), 6 samples tested negative, 4 tested inconclusive, and 58 tested positive.
- **Diagnostic Sensitivity:** 84.4 % (58/68)
Based on the diagnostic specificity and sensitivity results, **diagnostic concordance is 91.4 % (223/244).**

PERFORMANCE

Caution: The data presented are not representative of kit operating specifications but constitute experimental evidence of how kit performance is aligned with the manufacturer’s stated specifications.
Precision and Reproducibility

The precision and reproducibility of the ZENIT RA CARDIOLIPIN IgG kit assays were assessed using a protocol based on the guidelines provided by Clinical and Laboratory Standards Institute (CLSI) document EP5-A2.

**Precision** was calculated by analyzing the results for 20 replicates of five sera (one negative and four positive at different anti-cardiolipin IgG concentrations) run with two different reagent lots during the same experimental session.

The anti-cardiolipin IgG concentration found in the negative serum (N4) fell in the intervals from 0.3 to 2.5 GPL U/ml and from 0.0 to 3.2 GPL U/ml when tested with reagents lots no. 1 and no. 2, respectively.

The results obtained with the 4 positive sera are reported in the table below.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Reagents Lot No.</th>
<th>Mean Concentration (GPL U/ml)</th>
<th>SD</th>
<th>CV %</th>
</tr>
</thead>
<tbody>
<tr>
<td>P1</td>
<td>1</td>
<td>24.0</td>
<td>1.12</td>
<td>4.7</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>22.2</td>
<td>1.05</td>
<td>4.7</td>
</tr>
<tr>
<td>P2</td>
<td>1</td>
<td>82.8</td>
<td>2.23</td>
<td>2.7</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>69.2</td>
<td>1.67</td>
<td>2.4</td>
</tr>
<tr>
<td>P3</td>
<td>1</td>
<td>158.7</td>
<td>5.59</td>
<td>3.5</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>133.4</td>
<td>5.53</td>
<td>4.1</td>
</tr>
<tr>
<td>P4</td>
<td>1</td>
<td>341.8</td>
<td>16.53</td>
<td>4.8</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>286.1</td>
<td>17.88</td>
<td>6.2</td>
</tr>
</tbody>
</table>

**Reproducibility** was calculated by analyzing the results for four sera (one negative and three positive at different anti-cardiolipin IgG concentrations) assayed in single replicates in 30 different sessions, with two different reagent lots.

The anti-cardiolipin IgG concentration found in the negative serum (N4) fell in the interval from 0.0 to 4.3 GPL U/ml.

The results obtained with the 3 positive sera are reported in the table below.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Mean Concentration (GPL U/ml)</th>
<th>SD</th>
<th>CV %</th>
</tr>
</thead>
<tbody>
<tr>
<td>P1</td>
<td>23.4</td>
<td>2.15</td>
<td>9.2</td>
</tr>
<tr>
<td>P2</td>
<td>71.6</td>
<td>5.19</td>
<td>7.2</td>
</tr>
<tr>
<td>P3</td>
<td>153.2</td>
<td>16.82</td>
<td>11.0</td>
</tr>
</tbody>
</table>

**Linearity of Dilution**

The linearity of the ZENIT RA CARDIOLIPIN IgG kit dilutions was evaluated by following a protocol based on the guidelines provided by Clinical and Laboratory Standards Institute (CLSI) document EP6-A.
Scaled dilutions of 3 sera containing high concentrations of anti-cardiolipin IgG, diluted with the sample dilution solution, were assayed.

The results of this study are summarized in the table below.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Dilution Factor</th>
<th>Measured Concentration (GPL U/ml)</th>
<th>Expected Concentration (GPL U/ml)</th>
<th>Recovery %</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1</td>
<td>201.5</td>
<td>-</td>
<td>(100)</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>106.4</td>
<td>100.8</td>
<td>105.6</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>52.8</td>
<td>50.4</td>
<td>104.8</td>
</tr>
<tr>
<td></td>
<td>8</td>
<td>23.9</td>
<td>25.2</td>
<td>94.8</td>
</tr>
<tr>
<td>2</td>
<td>1</td>
<td>195.1</td>
<td>-</td>
<td>(100)</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>102.2</td>
<td>97.6</td>
<td>104.8</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>50.6</td>
<td>48.8</td>
<td>103.7</td>
</tr>
<tr>
<td></td>
<td>8</td>
<td>21.8</td>
<td>24.4</td>
<td>89.3</td>
</tr>
<tr>
<td>3</td>
<td>1</td>
<td>470.6</td>
<td>-</td>
<td>(100)</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>197.7</td>
<td>235.3</td>
<td>84.0</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>86.9</td>
<td>117.7</td>
<td>73.8</td>
</tr>
<tr>
<td></td>
<td>8</td>
<td>41.6</td>
<td>58.8</td>
<td>70.7</td>
</tr>
</tbody>
</table>

It must in any case be noted that not all sera, when measured at different dilutions, can give non-linear results within the measurability interval, since the result is dependent not only on concentration but also on the affinity of the antibodies in the sample.

**Analytical Sensitivity**

The analytical sensitivity of the ZENIT RA CARDIOLIPIN IgG kit, expressed as the Limit of Detection (LoD: the smallest quantity of analyte that can be measured by the method), was evaluated by following a protocol based on the guidelines provided by Clinical and Laboratory Standards Institute (CLSI) document EP17-A; the formula for calculating the limit of detection is $\text{LoD} = \text{LoB} + C_\beta SD_s$ (where $\text{LoB}$ is the value of the Limit of Blank, $SD_s$ is the estimated standard deviation of low-concentration sample distribution, and $C_\beta$ is derived from the 95th percentile of standard normal [Gaussian] distribution).

Four (4) samples at low analyte concentration were assayed in single replicates, using two different reagents lots, in 30 different experiments.

The resulting Limit of Detection of the ZENIT RA CARDIOLIPIN IgG kit was 10.8 GPL U/ml.

The Limit of Detection values, clinical considerations, and the results of comparison with reference methods contributed to definition of the cutoff value.

**Analytical Specificity: Interferences**

A study based on the guidelines provided by CLSI document EP7-A2 demonstrated that assay performance is not influenced by inclusion, in the sample, of the potentially interfering substances listed below at concentrations up to those tested.
Use of lipemic, hemolyzed, or turbid samples is not recommended.

**Analytical Specificity: Cross-reactivity**

A study of 26 samples, all with high levels of other autoantibodies and negative for anti-cardiolipin IgG, was conducted to evaluate potential cross-reactions with the antigen used for sensitizing the magnetic particles. The samples were so divided: SS-A (2), SS-B (3), U1-snRNP (1), Jo-1 (2), Scl-70 (3), Cenp B (2), histone (2), nucleolar (1), Gliadin/t-TG (3), CCP (1), RF (1), dsDNA (3), MPO (1), PR3 (1). The study revealed no significant cross-reactions between the solid-phase antigen and the other autoantibodies.

**High-dose Hook Effect**

Some methods for immunoassay of samples containing extremely high concentrations of analyte may provide apparent analyte levels that underestimate actual content (high-dose saturation or hook effect). The dual-incubation method employed by the ZENIT RA CARDIOLIPIN IgG kit is not influenced by this effect. A sample containing an extremely high concentration (above the top limit of the measurement interval) of anti-cardiolipin IgG has confirmed the absence of the hook effect up to a concentration of 815 GPL U/ml.

**Relative Sensitivity and Specificity**

The presence of anti-cardiolipin IgG antibodies was determined, using the ZENIT RA CARDIOLIPIN IgG kit and a commercially-available ELISA method, in 234 samples: 60 samples from patients affected with antiphospholipid antibody syndrome (APS), 45 samples from patients affected with systemic autoimmune rheumatic disorders, 26 samples from patients affected with various infectious diseases, and 99 samples from normal subjects. The ZENIT RA assay and the commercially-available ELISA assay gave discordant results for six (6) samples.

**Relative concordance** was therefore 97.4 % (228/234).

**Relative sensitivity** was shown to be 95.2 % (60/63).

**Relative specificity** was shown to be 98.2 % (168/171).
Calibration and Reference Preparations

The system was calibrated against the internationally-recognized “HARRIS” reference sera (Product no. 6495GM, Lot no. 021008).

The quantity of anti-cardiolipin IgG antibodies contained in the “Human IgG Anti-Cardiolipin Monoclonal Antibody EY2C9 1.1 µg/ml (CDC, Cat. No. IS2717, Lot no. 05-0058 09/20/05)” reference reagent, reconstituted with 0.5 ml (protein concentration: 1.1 µg/ml), tested with the ZENIT RA CARDIOLIPIN IgG kit, was 55.6 GPL U/ml.

BIBLIOGRAPHY


